



Total synthesis of a cholinergic neuron-specific ganglioside GT1a α : A high affinity ligand for myelin-associated glycoprotein (MAG)[†]

Hiroimi Ito¹, Hideharu Ishida¹, Hatsue Waki², Susum Ando² and Makoto Kiso^{1*}

¹Department of Applied Bioorganic Chemistry, Gifu University, Gifu 501-1193, Japan

²Department of Membrane Biochemistry, Tokyo Metropolitan Institute of Gerontology, 35-2 Sakae-cho, Itabashi-ku, Tokyo 173-0015, Japan

An efficient total synthesis of a cholinergic neuron-specific ganglioside GT1a α (IV³NeuAcIII⁶NeuAcII³NeuAc-GgOse4Cer) is described. The suitably protected sialyl- α (2 \rightarrow 6)-ganglioside (III⁶NeuAc-GgOse3) derivative was glycosylated with the phenyl 2-thioglycoside of sialic acid in the presence of N-iodosuccinimide (NIS) and trimethylsilyl trifluoromethanesulfonate (TMSOTf) in acetonitrile medium, giving the disialoganglioside (III⁶NeuAcII³NeuAc-GgOse3) derivative which contains both sialyl- α (2 \rightarrow 6)-GalNAc and sialyl- α (2 \rightarrow 3)-Gal structures (Route I). This pentasaccharide was efficiently synthesized also by the coupling of (methyl 5-acetamido-4,7,8,9-tetra-*O*-acetyl-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyranosylate)-(2 \rightarrow 6)-2-deoxy-3,4-*O*-isopropylidene-2-phthalimido-D-galactopyranosyl trichloroacetimidate with 2-(trimethylsilyl)ethyl (methyl 5-acetamido-4,7,8,9-tetra-*O*-acetyl-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyranosylate)-(2 \rightarrow 3)-(2,6-di-*O*-benzyl- β -D-galactopyranosyl)-(1 \rightarrow 4)-2,3,6-tri-*O*-benzyl- β -D-glucopyranoside, followed by conversion of the phthalimido group to the acetamido group (Route II). *O*-Deisopropylideneation and further glycosylation with methyl (methyl 5-acetamido-4,7,8,9-tetra-*O*-acetyl-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyranosylate)-(2 \rightarrow 3)-2,4,6-tri-*O*-benzoyl-1-thio- β -D-galactopyranoside, promoted by dimethyl(methylthio)sulfonium triflate (DMTST), gave the desired trisialoganglioside (IV³NeuAcIII⁶NeuAcII³NeuAc-GgOse4) derivative, which was converted stepwise into the title ganglioside GT1a α by the introduction of the ceramide part and then complete deprotection. The ganglioside obtained was shown to be identical with the native GT1a α on TLC-immunostaining.

Keywords: ganglioside GT1a α , sialic acid, cholinergic neuron-specific antigen, myelin-associated glycoprotein

Introduction

Gangliosides, sialic acid-containing glycosphingolipids, are a class of structurally diverse molecules commonly present in cell-surface membranes and are particularly rich in tissues of the central nervous system. It has been widely recognized that gangliosides are involved in many biological processes such as cell growth, cell differentiation, cell adhesion, immune responses, oncogenesis, and many other receptor-mediated reactions [2–5]. The sialo-oligosaccharide parts of gangliosides are considered to be exposed as ligands to the external environment, capable of expressing biological functions which are congruent with the chemical structure of the ganglioside.

A new class (α -series) of gangliosides, GT1a α and GQ1b α , have been identified [6–11] as cholinergic neuron-specific antigens in brains. These gangliosides have in common one sialic acid residue α (2 \rightarrow 6)-linked to the GalNAc moiety in the ganglioside core structure (Fig. 1). We have reported the first total syntheses of α -series gangliosides, GM1 α [12], GD1 α [13], and GQ1b α [14] to elucidate their biological functions. Recently, it has been found [15–17] that the synthetic GQ1b α has the extremely high potency as the ligand of myelin-associated glycoprotein (MAG), a member of sialic acid-dependent immunoglobulin lectin (siglec) family, previously termed sialoadhesins or I-type lectins [18–20]. The binding activity of GQ1b α to MAG was much higher than that of GT1b, suggesting a special importance of the sialic acid residue α (2 \rightarrow 6)-linked to the GalNAc moiety for the MAG binding. In view of these facts, we describe herein an efficient total synthesis of ganglioside GT1a α , a cholinergic neuron-specific antigen.

[†]Synthetic Studies on Sialoglycoconjugates, Part 112. For Part 111, see Ref. [1].

*Corresponding author. Tel and Fax: +81-58-293-2916/2918; E-mail: kiso@cc.gifu-u.ac.jp

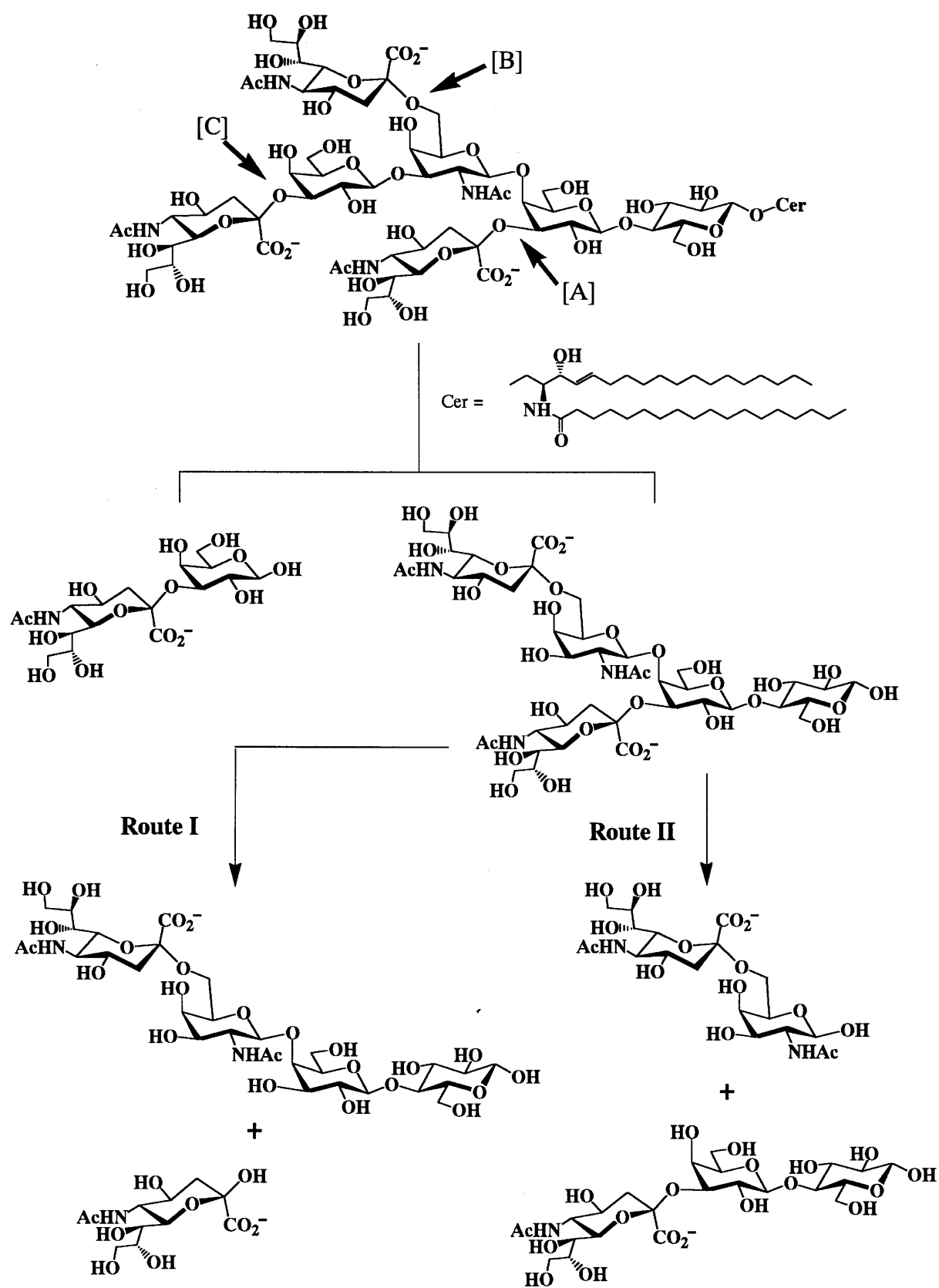


Figure 1. Retrosynthetic analysis of angioside GT1 α z

Results and discussion

The most important problems in the total synthesis of the title trisialoganglioside GT1ax are a) regio- and α -stereoselective sialylations at *O*-3 of both internal and external Gal residues (arrows [A] and [C] in Fig. 1), and at *O*-6 of the GalNAc residue (arrow [B] in Fig. 1), b) efficient constructions of the gangliotriose and gangliotetraose core structures, and c) convenient transformation of the protected heptasaccharide chain (IV³NeuAcIII⁶NcuAcII³NeuAc-GgOse4) into the target ganglioside. We employed two synthetic routes (I and II) to construct the key intermediate, III⁶NeuAcII³NeuAcGgOse3.

In the route I [21], we selected the suitably protected sialyl- α (2 \rightarrow 6)-gangliotriose derivative **1** as a key glycosyl acceptor, which had served as the synthetic intermediate for GQ1b α [14] and the phenyl 2-thioglycoside of sialic acid (**2**) as the sialyl donor [22] (Fig. 2). The glycosylations of **1** with **2** was performed in the presence of N-iodosuccinimide (NIS)-trimethylsilyl trifluoromethanesulfonate (TMSOTf) [22,25] in acetonitrile medium [22–29], to give **3** in 40% yield. Due to the low yield of the above mentioned reaction, an alternative synthetic route, route II, was developed for the synthesis of GT1ax as shown in Figure 3.

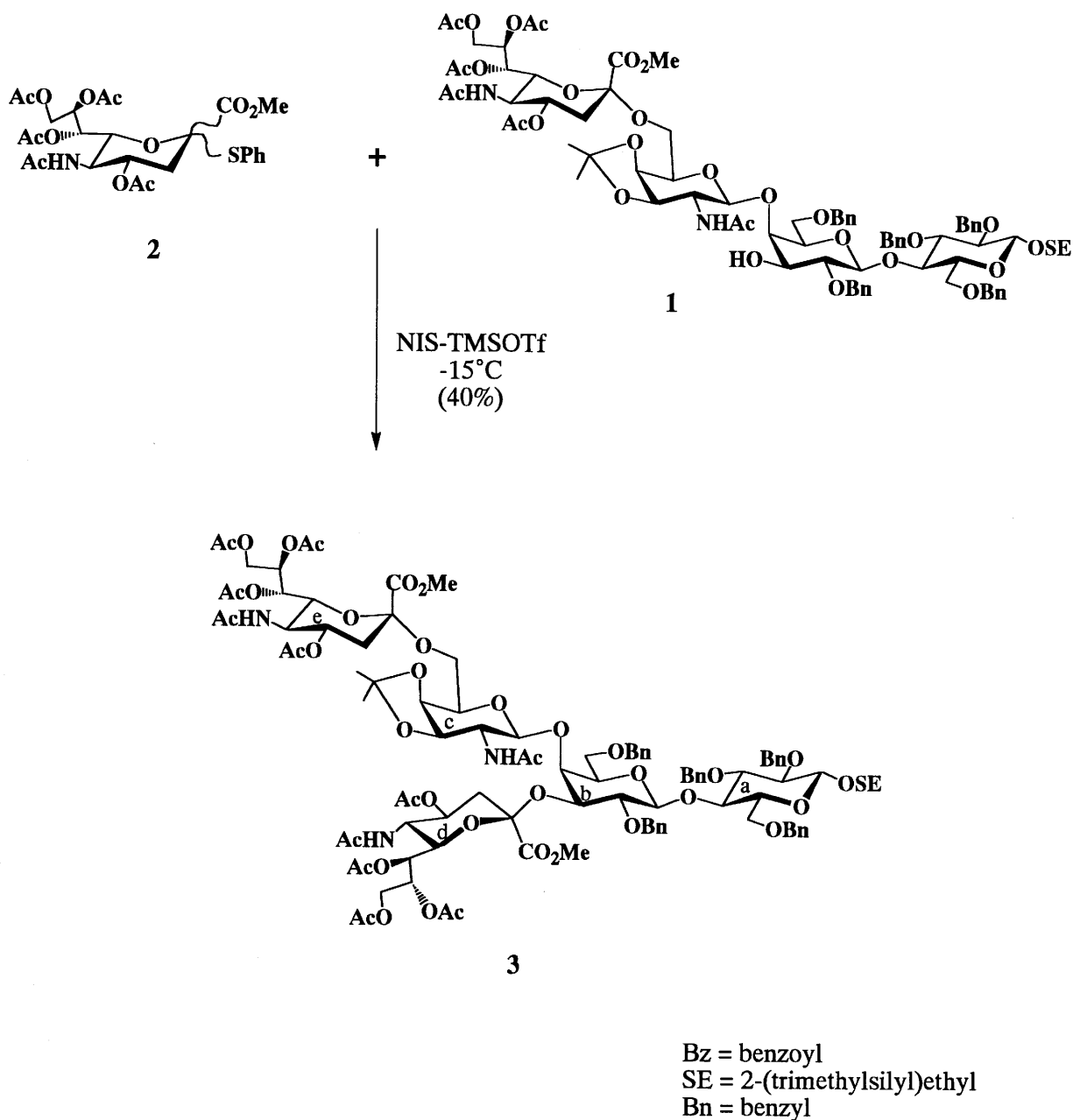


Figure 2. Synthesis of disialogangliotriose derivative by Route I

3,4,6-Tri-*O*-acetyl-2-deoxy-2-phthalimido-D-galactopyranose, which was readily prepared by selective 1-*O*-deacetylation of 1,3,4,6-tera-*O*-acetyl-2-deoxy-2-phthalimido-D-galactopyranose [28,32], was treated with *t*-butyldimethylsilyl (TBDMS) chloride and imidazole in *N,N*-dimethylformamide (DMF) to give **4** in 92% yield. The characteristic $^1\text{H-NMR}$ signal at δ 5.48 (d, $J_{1,2} = 8.1$ Hz, H-1) showed the anomeric configuration of **4** to be β . *O*-Deacetylation of **4** and the following 3,4-*O*-isopropylideneation gave **6**, which was glycosylated with NeuAc donor **2** in the presence of NIS-TfOH in acetonitrile at -30°C to afford the desired sialyl- $\alpha(2\rightarrow6)$ glycoside **7** in 81% yield; no β -glycoside was isolated. In the $^1\text{H-NMR}$ spectrum of **7**, the characteristic H-3 ax (δ 1.98, t, $J_{\text{gem}} = J_{3ax,4} = 12.8$ Hz), H-3 eq (δ 2.63, dd, $J_{\text{gem}} = 12.8$, $J_{3eq,4} = 4.6$ Hz) and a three-proton singlet of MeO (δ 3.84) of the NeuAc residue were clearly observed indicating the structure assigned. Removal of the TBDMS group in **7** (83%) and trichloroacetimidate formation (99%) yielded the disaccharide donor **9** (the major compound was β -imidate, $J_{1,2} = 9.2$ Hz), which was coupled with the sialyl- $\alpha(2\rightarrow3)$ lactose derivative **10** [28] to give the desired pentasaccharide (**11**, 76%). The characteristic one-proton doublet of H-1c (δ 5.22, $J_{1,2} = 8.7$ Hz) in the $^1\text{H-NMR}$ spectrum of **11** indicated the newly formed glycosidic linkage to be β . The phthalimido group in **11** was then converted to the acetamido group. The physicochemical properties and spectral data of **3** thus obtained were identical with those of **3** separately synthesized by the coupling of **1** and **2** as described above. In the $^1\text{H-NMR}$ spectrum of **3**, H-3 eq of the NeuAc $\alpha(2\rightarrow6)$ -linked to the GalNAc residue was observed at δ 2.61 (dd, $J_{\text{gem}} = 12.6$, $J_{3eq,4} = 4.6$ Hz), while H-3 eq of the NeuAc $\alpha(2\rightarrow3)$ -linked to the Gal residue at δ 2.23 showing significant up-field shift from δ 2.47 observed for **10**. The two, three-proton singlets of MeO appeared at δ 3.83 and δ 3.85, respectively.

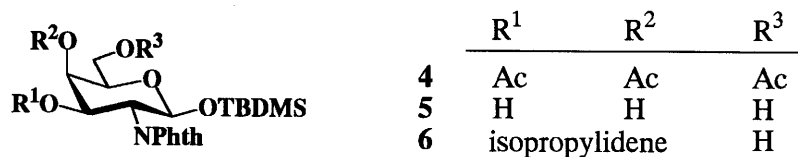
The isopropylidene group in **3** was cleaved by treatment with 80% acetic acid for 3 h at 40°C to give **13** in 85% yield. Regioselective glycosylation of **13** with methyl (methyl 5-acetamido-4,7,8,9-tetra-*O*-acetyl-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyranosylonate)-(2 \rightarrow 3)-2,4,6-tri-*O*-benzoyl-1-thio- β -D-galactopyranoside (**14**) [33] in dichloromethane for 24 h at 0°C in the presence of DMTST and molecular sieves 4 Å (MS-4 Å), gave the protected GT1 α heptasaccharide **15** in 95% yield (Fig. 4). Hydrogenolytic removal of the benzyl groups in **15** over Pd(OH) $_2$ in 9:1 ethanol-acetic acid, followed by complete acetylation of the resulting free hydroxyl groups with Ac $_2$ O-pyridine, afforded the fully acylated oligosaccharide **16** in 81% yield. Selective removal of the 2-(trimethylsilyl)ethyl (SE) group was achieved [34,35] by treatment of **16** with trifluoroacetic acid in dichloromethane to give the 1-hydroxy compound **17** (92%), which upon further treatment [35] with trichloroacetonitrile in the presence of 1,8-diazabicyclo[5.4.0]undec-7-ene (DBU) in dichloromethane, gave the trichloroacetimidate **18** in high yield.

Coupling of **18** with (2*S*,3*R*,4*E*)-2-azido-3-*O*-(*tert*-butyldiphenylsilyl)-4-octadecanamido-1,3-diol (**19**) [36,37] was car-

ried out in the presence of TMSOTf and MS-4 Å (AW300) in dichloromethane to give **20** in 62% yield. Selective reduction [38] of the azido group in **20** with triphenylphosphine in 5:1 benzene-water gave the amine, which on condensation with stearic acid using 1-(3-dimethylamino-propyl)-3-ethylcarbodiimide hydrochloride (WSC) in dichloromethane, afforded the fully protected ganglioside GT1 α **21** in 76% yield. Finally, removal [36] of the *tert*-butyldiphenylsilyl group in **21** with 1.0 M tetrabutylammonium fluoride in acetonitrile, *O*-deacetylation with sodium methoxide in methanol, and subsequent saponification of the methyl ester group gave the title ganglioside GT1 α as amorphous mass in quantitative yield, after chromatography on a column of Sephadex LH-20 with 5:5:1 chloroform-methanol-water.

The structure of ganglioside GT1 α thus obtained was characterized by FAB MS (negative ion mode) and $^1\text{H-NMR}$ [6] spectrometry. The molecular ion species of the synthetic GT1 α were clearly detected at m/z 2172.04 [M-Na] $^-$ (C $_{95}$ H $_{162}$ N $_5$ Na $_2$ O $_{47}$ MW, Exact 2171.0236, Ave. 2172.3161), 2150.06, 2149.07 [M-2Na] $^-$, and 2127.06 [M-3Na] $^-$, accompanied by the significant fragment ions at m/z 1858.0, 1857.0 [M-2Na-NeuAc] $^-$, 1696.0 [M-2Na-NeuAc-Gal] $^-$, and 1545.0 [M-3Na-2NeuAc] $^-$; providing unambiguous evidences for the structure assigned. The characteristic fragment ions at m/z 888.7 [lactosylceramide] $^-$, 726.6 [glucosylceramide] $^-$, and 564.6 [ceramide] $^-$, provides further evidences for the assigned structure including the ceramide moiety. In the $^1\text{H-NMR}$ spectrum (500 MHz) of the synthetic GT1 α in DMSO- d_6 -D $_2$ O, the characteristic three H-3 eq of NeuAc (δ 2.73–2.89), four anomeric protons due to the β glycosidic linkages in the GgOse4 core structure (δ 4.15, 4.30, 4.43 and 4.84), and two olefinic protons of ceramide (δ 5.32 and 5.75) were clearly observed. In addition, TLC-immunostaining of the synthetic GT1 α was performed in comparison with the native GT1 α . As shown in Figure 6, the two compounds were found to be identical.

MAG is a quantitatively minor protein constituent of central and peripheral nervous system myelin, and has been considered to be implicated in myelin-neuron interactions [39]. Therefore, detailed structure-function relationship study for carbohydrate ligands that support MAG binding may provide opportunities for intervention in the control of neurite outgrowth and myelination. In binding experiments with the MAG-transfected COS cell for the immobilized gangliosides including synthetic GT1 α , it was demonstrated that GT1 α can efficiently support the adhesion as strong as GD1 α [1]. The hierarchy of binding strengths of a series of gangliosides with MAG were GQ1b α > GT1 α , GD1 α > GT1b, GD1a > GM3, GM4 (GQ1b, GD1b, GM1 and GD3 failed to support binding) [39,40]. This result indicates that not only the terminal sialic acid, which is essential for MAG binding [15–17], but also the internal sialic acids play important roles for expressing higher binding activity for MAG.



TBDMS = *tert*-butyldimethylsilyl
 NPhth = phthalimido

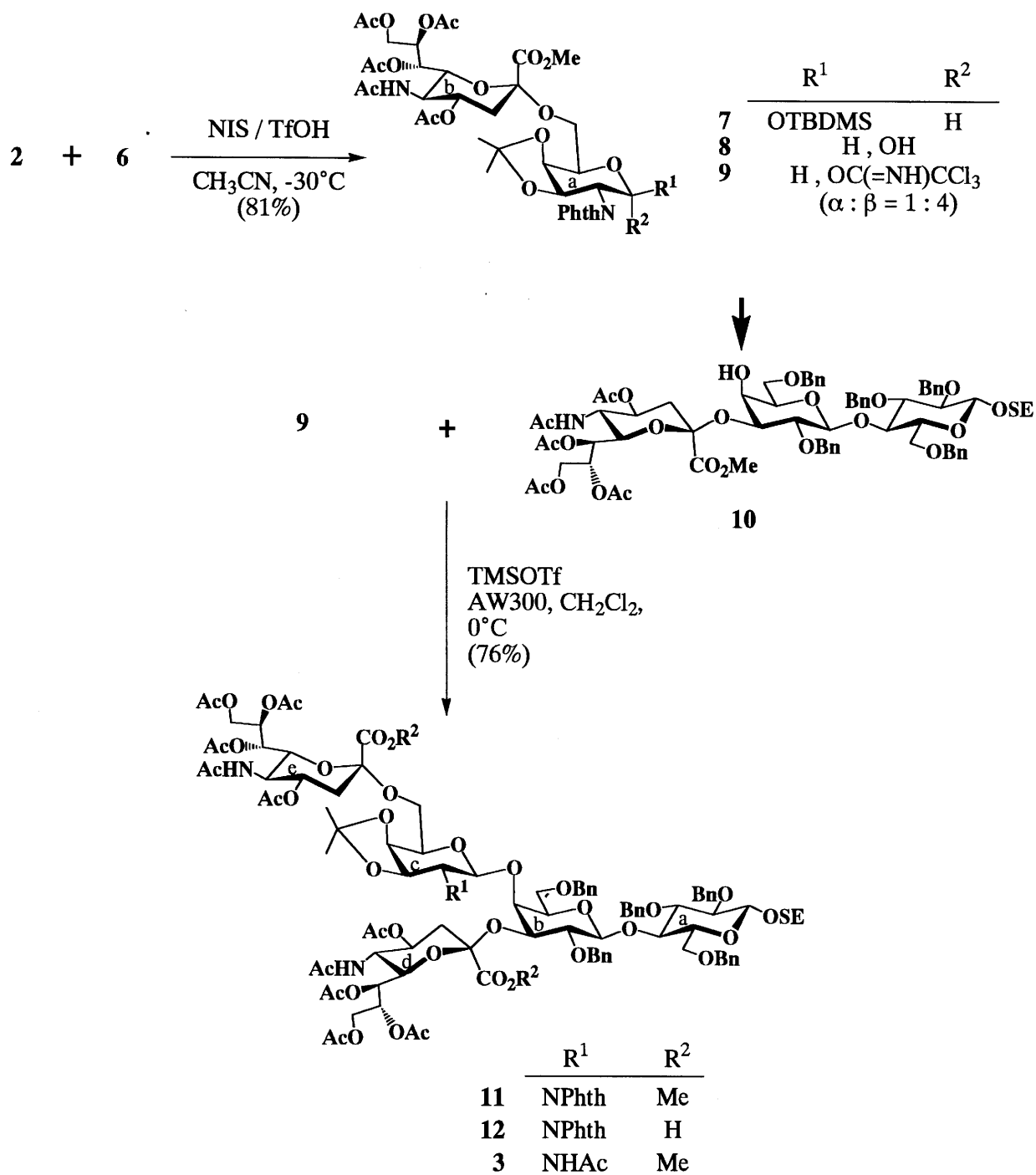


Figure 3. Synthesis of disialoganglioside derivative by Route II

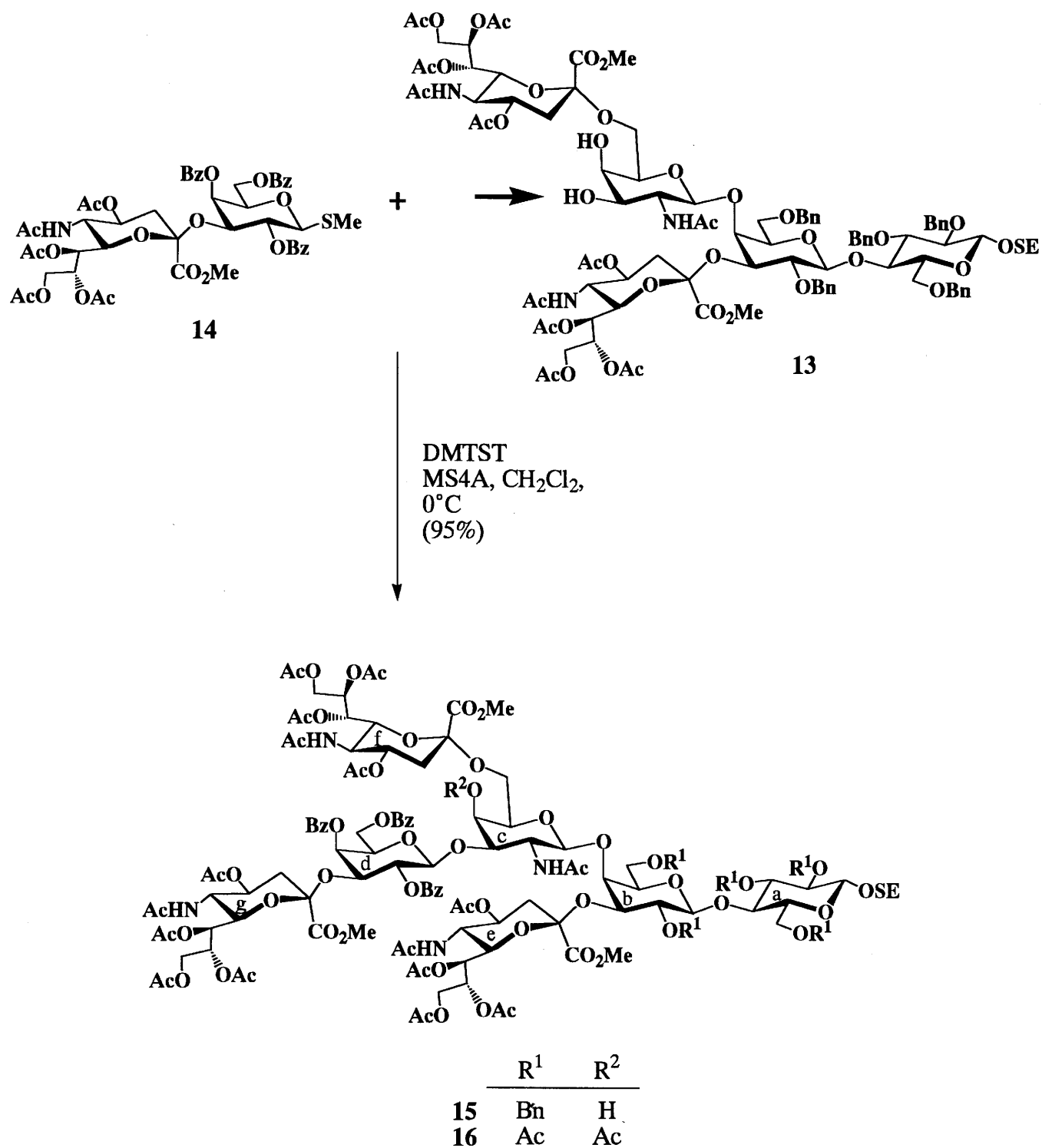
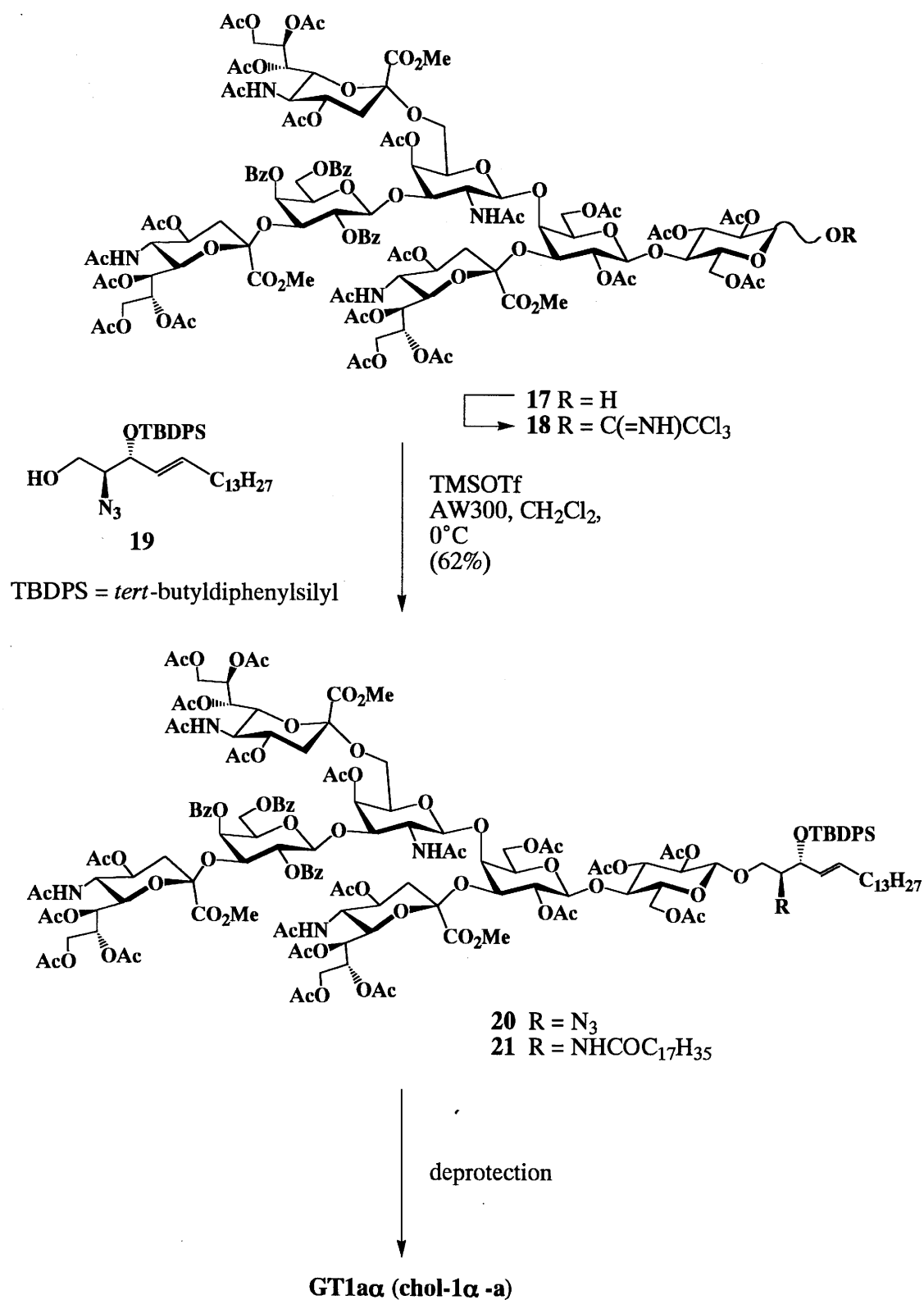


Figure 4. Synthesis of trisialogangliotetraose derivative

Figure 5. Total synthesis of ganglioside GT1a α

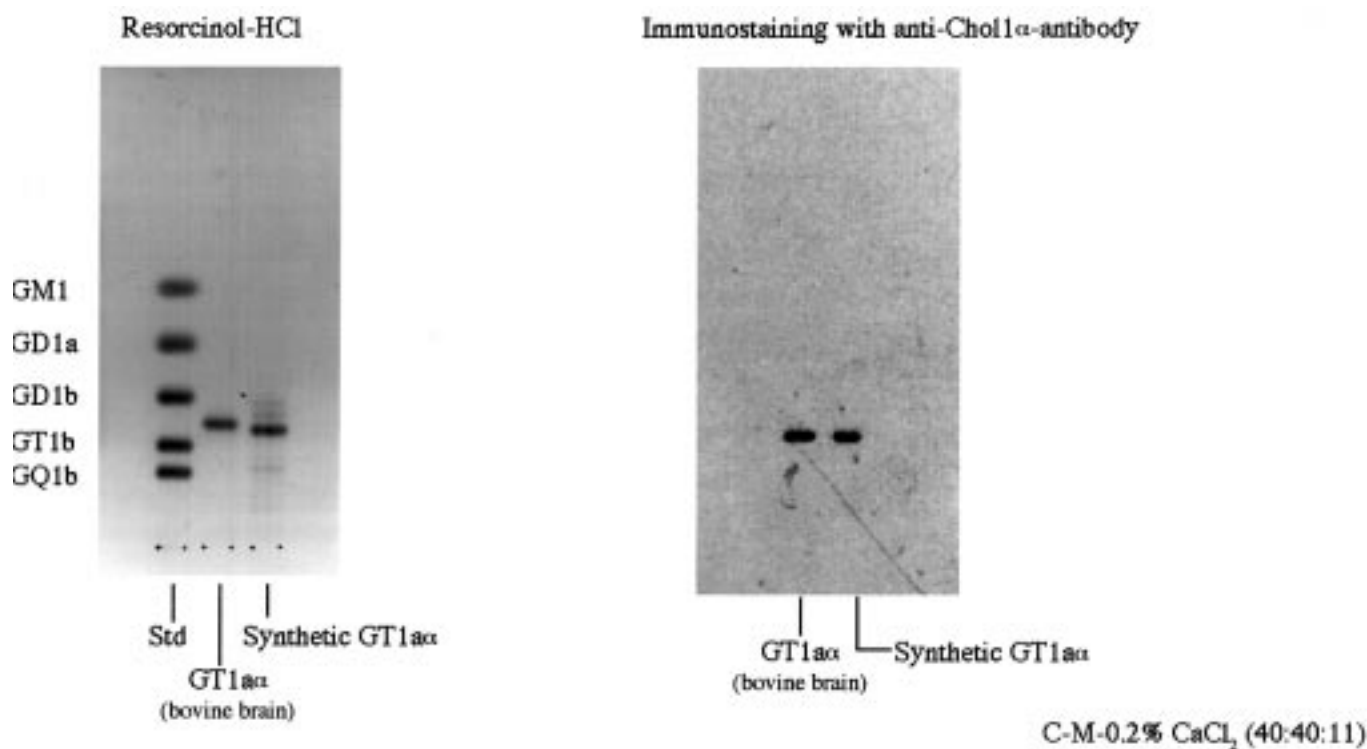


Figure 6. TLC-immunostaining of synthetic and native GT1 α x

Materials and methods

Chemical synthesis

General methods

Optical rotations were determined with a Union PM-201 polarimeter at 25°C and IR spectra were recorded with a Jasco IRA-100 spectrophotometer. ¹H-NMR spectra were recorded at 400 or 500 MHz with a Varian UNITY Inova-400 and a Varian UNITY Inova-500 spectrometers, respectively. FAB-MS were recorded on a Jeol JMS-SX 120 A mass spectrometer/JMA-DA 7000 data system. Preparative chromatography was performed on silica gel (Fuji Silysia Co., 300 mesh) with the solvent systems specified. Concentrations were conducted in vacuo.

2-(Trimethylsilyl)ethyl *O*-(methyl 5-acetamido-4,7,8,9-tetra-*O*-acetyl-3,5-dideoxy-*D*-glycero- α -*D*-galacto-2-nonulopyranosylonate)-(2 \rightarrow 6)-*O*-(2-acetamido-2-deoxy-3,4-*O*-isopropylidene- β -*D*-galactopyranosyl)-(1 \rightarrow 4)-*O*-[(methyl 5-acetamido-4,7,8,9-tetra-*O*-acetyl-3,5-dideoxy-*D*-glycero- α -*D*-galacto-2-nonulopyranosylonate)-(2 \rightarrow 3)]-*O*-(2,6-di-*O*-benzyl- β -*D*-galactopyranosyl)-(1 \rightarrow 4)-2,3,6-tri-*O*-benzyl- β -*D*-glucopyranoside (**3**)

Route (I):

To a solution of 2-(trimethylsilyl)ethyl *O*-(methyl 5-acetamido-4,7,8,9-tetra-*O*-acetyl-3,5-dideoxy-*D*-glycero- α -*D*-galacto-

2-nonulopyranosylonate)-(2 \rightarrow 6)-*O*-(2-acetamido-2-deoxy-3,4-*O*-isopropylidene- β -*D*-galactopyranosyl)-(1 \rightarrow 4)-*O*-(2,6-di-*O*-benzyl- β -*D*-galactopyranosyl)-(1 \rightarrow 4)-*O*-2,3,6-tri-*O*-benzyl- β -*D*-glucopyranoside [**14**] (**1**; 800 mg, 0.50 mmol) and methyl (phenyl 5-acetamido-4,7,8,9-tetra-*O*-acetyl-3,5-dideoxy-2-thio-*D*-glycero-*D*-galacto-2-nonulopyranosid)onate [**30**] (**2**; 580 mg, 1.0 mmol) in acetonitrile (6 mL) were added molecular sieves 3 Å (1 g) and the mixture was stirred for 7 h at room temperature, then cooled to -20°C. To the solution were added *N*-iodosuccinimide (NIS; 675 mg, 3.0 mmol) and trimethylsilyl trifluoromethanesulfonate (TMSOTf; 58 μ L, 0.3 mmol), and the stirring was continued for 72 h at -15°C. After reaction completed, the precipitates were filtered off and washed with chloroform. The filtrate and washings were combined, and the solution was successively washed with M Na₂CO₃ and M Na₂S₂O₃, dried (Na₂SO₄) and concentrated. Column chromatography (20:1 toluene-methanol) of the residue on silica gel gave **3** (450 mg, 40%) as an amorphous mass.

Route (II):

To a solution of **12** (500 mg, 0.23 mmol) in aq. 95% ethanol (15 mL) was added hydrazine monohydrate (130 μ L, 2.3 mmol), and the solution was stirred for 15 h at 80°C. The solids were filtered and washed with ethanol, and the combined filtrate and washings was concentrated to a syrup. To a solution of the residue in methanol (10 mL) was added acetic anhydride (0.3 mL), and the solution was stirred for 6 h

at room temperature. Pyridine (0.6 mL) was added to the solution, and which was concentrated. To a solution of the residue in methanol (10 mL) was added large excess of diazomethane in diethyl ether (3 mL) at room temperature. After completion of the reaction, acetic acid (1 mL) was added to the solution, and which was concentrated and acetylated with pyridine (6 mL) and acetic anhydride (3 mL) for 24 h at 40°C. The product was purified by column chromatography (30:1 chloroform-methanol) of the syrup on silica gel to give **3** (344 mg, 71%) as an amorphous mass: $[\alpha]_D + 2.77^\circ$ (*c* 0.7, CHCl₃); IR (KBr) 3300 (NH), 3100–2900 (CH), 1750 and 1220 (ester), 1660 and 1530 (amide), 860 and 840 (Me₃Si), and 700 cm⁻¹ (phenyl); ¹H-NMR (CDCl₃) δ 1.03 (m, 2H, Me₃SiCH₂CH₂), 1.34 and 1.50 (2s, 6H, Me₂C), 1.66–2.12 (11s, 33H, AcN and 10AcO), 2.23 (m, 1H, H-3_{deq}), 2.61 (dd, 1H, *J*_{gem} = 12.6 Hz, *J*_{3eq,4} = 4.6 Hz, H-3_{eeq}), 3.83 and 3.85 (2s, 6H, 2MeO), 4.86 (m, 1H, H-4e), 5.10 (m, 1H, H-4d), and 7.16–7.57 (m, 25H, 5Ph).

Anal. Calcd for C₁₀₃H₁₃₅N₃O₄₀Si (2083.28): C, 59.38; H, 6.53; N, 2.02. Found: C, 59.37; H, 6.27; N, 2.01.

t-Butyldimethylsilyl 3,4,6-tri-*O*-acetyl-2-deoxy-2-phthalimido- β -D-galactopyranoside (**4**)

To a solution of 3,4,6-tri-*O*-acetyl-2-deoxy-2-phthalimido- β -D-galactopyranose (3.30 g, 7.6 mmol) in *N,N*-dimethylformamide (DMF; 10 mL) were added *t*-butyldimethylsilyl chloride (1.37 g, 9.1 mmol) and imidazole (1.24 g, 18.2 mmol) at 0°C. The mixture was stirred for 4 h at room temperature, and methanol (5 mL) was then added. The solution was concentrated to a syrup, and which was extracted with ethyl acetate. The organic layer was successively washed with water, dried (Na₂SO₄) and concentrated. Column chromatography (1:3 ethyl acetate-hexane) of the residue on silica gel gave **4** (3.82 g, 92%) as an amorphous mass: $[\alpha]_D - 12.66^\circ$ (*c* 0.6, CHCl₃); IR (KBr) 3000–2870 (CH), 1730 and 1240 (ester), 1700 (imide), and 720 cm⁻¹ (Ph); ¹H-NMR (CDCl₃) δ -0.05 and 0.05 (2s, 6H, SiMe₂), 0.70 (s, 9H, SiCMe₃), 1.86, 2.06 and 2.21 (3s, 9H, 3AcO), 4.09 (m, 1H, H-5), 4.15 (dd, 1H, *J*_{gem} = 11.0 Hz, H-6), 4.23 (dd, 1H, *J*_{gem} = 11.0 Hz, H-6'), 4.50 (dd, 1H, *J*_{1,2} = 8.1 Hz, *J*_{2,3} = 11.7 Hz, H-2), 5.48 (d, 1H, *J*_{1,2} = 8.1 Hz, H-1), 5.49 (d, 1H, H-4), 5.85 (dd, 1H, *J*_{2,3} = 11.7 Hz, *J*_{3,4} = 3.7 Hz, H-3), and 7.74–7.88 (m, 4H, Ph).

Anal. Calcd for C₂₆H₃₅NO₁₀Si (549.65): C, 56.82; H, 6.42; N, 2.55. Found: C, 56.81; H, 6.33; N, 2.44.

t-Butyldimethylsilyl 2-deoxy-2-phthalimido- β -D-galactopyranoside (**5**)

To a solution of **4** (3.80 g, 6.9 mmol) in 3:2 methanol-tetrahydrofuran (25 mL) was added sodium methoxide

(20 mg), and the mixture was stirred for 2 h at room temperature, then neutralized with Amberlite IR-120 (H⁺) resin. The resin was filtered and washed with methanol, and the combined filtrate and washings was concentrated. Column chromatography (10:1 chloroform-methanol) of the residue on silica gel gave **5** (2.75 g, 94%) as an amorphous mass: $[\alpha]_D - 9.35^\circ$ (*c* 0.6, CHCl₃); IR (KBr) 3500–2850 (OH, CH), 1700 (imide), and 710 cm⁻¹ (Ph); ¹H-NMR (CDCl₃) δ -0.09 and 0.04 (2s, 6H, SiMe₂), 0.68 (s, 9H, SiCMe₃), 3.69 (m, 1H, H-5), 3.89–3.93 (m, 2H, H-6,6'), 4.16 (d, 1H, *J*_{3,4} = 2.9 Hz, H-4), 4.30 (dd, 1H, *J*_{1,2} = 8.1 Hz, *J*_{2,3} = 11.0 Hz, H-2), 4.45 (dd, 1H, *J*_{2,3} = 11.0 Hz, *J*_{3,4} = 2.9 Hz, H-3), 5.36 (d, 1H, *J*_{1,2} = 8.1 Hz, H-1), and 7.70–7.82 (m, 4H, Ph).

Anal. Calcd for C₂₀H₂₉NO₇Si (423.54): C, 56.72; H, 6.90; N, 3.31. Found: C, 56.43; H, 6.89; N, 3.20.

t-Butyldimethylsilyl 2-deoxy-3,4-*O*-isopropylidene-2-phthalimido- β -D-galactopyranoside (**6**)

To a solution of **5** (3.70 g, 8.7 mmol) in DMF (20 mL) was added 2,2-dimethoxypropane (2.6 mL, 21.7 mmol) and drierite (4.0 g), and the mixture was stirred for 1 h at room temperature. After addition of (\pm)-10-Camphorsulfonic acid (100 mg), the mixture was stirred for 4 h at 80°C, then neutralized with triethylamine and concentrated. Column chromatography (1:2 ethyl acetate-hexane) of the residue on silica gel gave **6** (3.03 g, 75%) as an amorphous mass: $[\alpha]_D + 14.58^\circ$ (*c* 0.5, CHCl₃); IR (KBr) 3600–3400 (OH), 3050–2850 (CH), 1700 (imide), and 720 cm⁻¹ (Ph); ¹H-NMR (CDCl₃) δ -0.10 and 0.03 (2s, 6H, SiMe₂), 0.64 (s, 9H, SiCMe₃), 1.29 and 1.61 (2s, 6H, CMe₂), 3.84 (m, 1H, H-5), 3.98–4.03 (m, 2H, H-6, 6'), 4.19 (d, 1H, *J*_{3,4} = 5.0 Hz, H-4), 4.21 (dd, 1H, H-2), 4.83 (dd, 1H, *J*_{2,3} = 9.4 Hz, *J*_{3,4} = 5.0 Hz, H-3), 5.28 (d, 1H, *J*_{1,2} = 8.5 Hz, H-1), and 7.68–7.81 (m, 4H, Ph).

Anal. Calcd for C₂₃H₃₃NO₇Si (463.60): C, 59.59; H, 7.18; N, 3.02. Found: C, 59.43; H, 6.98; N, 2.94.

t-Butyldimethylsilyl *O*-(methyl 5-acetamido-4,7,8,9-tetra-*O*-acetyl-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyranosylonate)-(2 \rightarrow 6)-2-deoxy-3,4-*O*-isopropylidene-2-phthalimido- β -D-galactopyranoside (**7**)

To a solution of **6** (1.20 g, 2.6 mmol) and **2** (3.02 g, 5.2 mmol) in acetonitrile (15 mL) were added molecular sieves 3 Å (4.0 g) and the mixture was stirred for 5 h at room temperature, then cooled to -30°C. To the mixture were added, with stirring, NIS (2.33 g, 10.4 mmol) and TfOH (92 μ L, 1.0 mmol), and the stirring was continued for 24 h at -30°C. The solids were removed by filtration and washed with chloroform. The combined filtrate and washings was successively washed with

M Na₂CO₃ and M Na₂S₂O₃, dried (Na₂SO₄) and concentrated. Column chromatography (30:1 toluene–methanol) of the residue on silica gel gave **7** (1.95 g, 81%) as an amorphous mass: [α]_D +4.73° (*c* 0.8, CHCl₃); IR (KBr) 3500–2900 (NH, CH), 1740 and 1240 (ester), 1700 (imide), 1650 and 1550 (amide), and 740 cm⁻¹ (Ph); ¹H-NMR (CDCl₃) δ = 0.04 and 0.08 (2s, 6H, SiMe₂), 0.66 (s, 9H, SiCMe₃), 1.33 and 1.64 (2s, 6H, CMe₂), 1.89, 2.04, 2.05, 2.15 and 2.16 (5s, 15H, AcN and 4AcO), 1.98 (t, 1H, $J_{\text{gem}} = J_{3\text{ax},4} = 12.8$ Hz, H-3 β ax), 2.63 (dd, 1H, $J_{\text{gem}} = 12.8$ Hz, $J_{3\text{eq},4} = 4.6$ Hz, H-3 β eq), 3.84 (s, 3H, MeO), 4.08 (m, 1H, H-5b), 4.19 (dd, 1H, $J_{\text{gem}} = 12.4$ Hz, H-9b), 4.22 (dd, 1H, $J_{1,2} = 8.5$ Hz, $J_{2,3} = 9.4$ Hz, H-2a), 4.37 (dd, 1H, $J_{\text{gem}} = 12.4$ Hz, H-9'b), 4.81 (dd, 1H, $J_{2,3} = 9.4$ Hz, $J_{3,4} = 5.0$ Hz, H-3a), 4.91 (m, 1H, H-4b), 5.27 (d, 1H, $J_{1,2} = 8.5$ Hz, H-1a), 5.34 (m, 1H, H-7b), 5.40 (m, 1H, H-8b), and 7.69–7.82 (m, 4H, Ph).

Anal. Calcd for C₄₃H₆₀N₂O₁₉Si (937.04): C, 55.12; H, 6.45; N, 2.99. Found: C, 54.85; H, 6.36; N, 2.97.

O-(Methyl 5-acetamido-4,7,8,9-tetra-*O*-acetyl-3,5-dideoxy-*D*-glycero- α -*D*-galacto-2-nonulopyranosylonate)-(2→6)-2-deoxy-3,4-*O*-isopropylidene-2-phthalimido- β -*D*-galactopyranosyl trichloroacetimidate (**9**)

To a solution of **7** (1.90 g, 2.0 mmol) in tetrahydrofuran (15 mL) were added acetic acid (150 μ L) and M tetrabutylammonium fluoride in tetrahydrofuran (3 mL) at -15°C, and the mixture was stirred for 17 h at -15°C. The mixture was diluted with chloroform and the solution was washed with water, dried (Na₂SO₄) and concentrated. Column chromatography (20:1 chloroform–methanol) of the residue on silica gel gave (methyl 5-acetamido-4,7,8,9-tetra-*O*-acetyl-3,5-dideoxy-*D*-glycero- α -*D*-galacto-2-nonulopyranosylonate)-(2→6)-2-deoxy-3,4-*O*-isopropylidene-2-phthalimido- β -*D*-galactopyranose (**8**; 1.38 g, 83%) as an amorphous mass: IR (KBr) 3600–3100 (OH, NH), 1730 and 1240 (ester), 1700 (imide), 1660 and 1550 (amide), and 720 cm⁻¹ (Ph). The ¹H-NMR data showed the complete loss of the TBDMS group at *O*-1.

To a solution of **8** (560 mg, 0.068 mmol) in dichloromethane (1.5 mL) and trichloroacetonitrile (3 mL, 2.0 mmol) was added 1,8-diazabicyclo[5.4.0]undec-7-ene (DBU; 120 μ L, 0.082 mmol) at 0°C, and the mixture was stirred for 30 min at 0°C, then concentrated. Column chromatography (35:1 chloroform–methanol) of the residue on silica gel gave **9** (656 mg, 99%, α : β = 1:4) as an amorphous mass: [α]_D +25.77° (*c* 0.5, CHCl₃); IR (KBr) 3400 (NH), 1720 and 1220 (ester), 1700 (imide), 1680 and 1550 (amide), and 720 cm⁻¹ (Ph); ¹H-NMR for β isomer (CDCl₃) δ 1.36 and 1.67 (2s, 6H, CMe₂), 1.90, 2.03, 2.04, 2.14 and 2.17 (5s, 15H, AcN and 4AcO), 2.00 (t, 1H, $J_{\text{gem}} = J_{3\text{ax},4} = 12.8$ Hz, H-3 β ax), 2.64 (dd, 1H, $J_{\text{gem}} = 12.8$ Hz, $J_{3\text{eq},4} = 4.6$ Hz, H-3 β eq), 3.83 (s,

3H, MeO), 4.90 (m, 1H, H-4b), 5.33 (m, 1H, H-7b), 5.49 (m, 1H, H-8b), 6.36 (d, 1H, $J_{1,2} = 9.2$ Hz, H-1a), 7.69–7.82 (m, 4H, Ph), and 8.60 (s, 1H, C = NH).

Anal. Calcd for C₃₉H₄₆Cl₃N₃O₁₉Si (967.16): C, 48.43; H, 4.79; N, 4.34. Found: C, 48.27; H, 4.55; N, 4.14.

2-(Trimethylsilyl)ethyl *O*-(methyl 5-acetamido-4,7,8,9-tetra-*O*-acetyl-3,5-dideoxy-*D*-glycero- α -*D*-galacto-2-nonulopyranosylonate)-(2→6)-*O*-(2-deoxy-3,4-*O*-isopropylidene-2-phthalimido- β -*D*-galactopyranosyl)-(1→4)-*O*-[(methyl 5-acetamido-4,7,8,9-tetra-*O*-acetyl-3,5-dideoxy-*D*-glycero- α -*D*-galacto-2-nonulopyranosylonate)-(2→3)]-*O*-(2,6-di-*O*-benzyl- β -*D*-galactopyranosyl)-(1→4)-2,3,6-tri-*O*-benzyl- β -*D*-glucopyranoside (**11**)

To a solution of 2-(trimethylsilyl)ethyl *O*-(methyl 5-acetamido-4,7,8,9-tetra-*O*-acetyl-3,5-dideoxy-*D*-glycero- α -*D*-galacto-2-nonulopyranosylonate)-(2→3)-*O*-(2,6-di-*O*-benzyl- β -*D*-galactopyranosyl)-(1→4)-2,3,6-tri-*O*-benzyl- β -*D*-glucopyranoside (**10**; 612 mg, 0.45 mmol) and **9** (656 mg, 0.68 mmol) in dichloromethane (5 mL) was added molecular sieves 4 Å (AW-300; 1.0 g), and the mixture was stirred for 15 h at room temperature, then cooled to 0°C. TMSOTf (13 μ L, 0.068 mmol) was added, and the mixture was stirred for 24 h at 0°C, then filtered. The insoluble materials were washed with M NaHCO₃ and water, and the combined filtrate and washings was dried (Na₂SO₄) and concentrated. Column chromatography (25:1 chloroform–methanol) of the residue on silica gel gave **11** (734 mg, 76%) as an amorphous mass: [α]_D +16.92° (*c* 0.5, CHCl₃); IR (KBr) 3500–3200 (NH), 1750 and 1230 (ester), 1700 (imide), 1670 and 1550 (amide), 860 and 840 (Me₃Si), and 700 cm⁻¹ (Ph); ¹H-NMR (CDCl₃) δ 1.02 (m, 2H, Me₃SiCH₂CH₂), 1.34 and 1.48 (2s, 6H, CMe₂), 1.68, 1.85, 1.86, 1.92, 2.01, 2.02, 2.03, 2.08, 2.09 and 2.11 (10s, 30H, 2AcN and 8AcO), 1.78 (t, 1H, $J_{\text{gem}} = J_{3\text{ax},4} = 12.8$ Hz, H-3 β dax), 1.88 (t, 1H, $J_{\text{gem}} = J_{3\text{ax},4} = 12.8$ Hz, H-3 β eax), 2.55 (dd, 1H, $J_{\text{gem}} = 12.8$ Hz, $J_{3\text{eq},4} = 4.8$ Hz, H-3 β eeq), 2.84 (dd, 1H, $J_{1,2} = 7.3$ Hz, $J_{2,3} = 9.6$ Hz, H-2b), 2.88 (dd, 1H, $J_{\text{gem}} = 12.8$ Hz, $J_{3\text{eq},4} = 4.3$ Hz, H-3 β deq), 3.31 (dd, 1H, $J_{1,2} = 7.8$ Hz, $J_{2,3} = 9.2$ Hz, H-2a), 3.53 (m, 1H, H-3a), 3.62 (d, 1H, $J_{3,4} = 2.3$ Hz, H-4b), 3.76 (t, 1H, $J_{3,4} = J_{4,5} = 9.5$ Hz, H-4a), 3.82 and 3.87 (2s, 6H, 2MeO), 4.03 (m, 2H, H-5d and H-5e), 4.10 (m, 1H, H-3b), 4.28 (dd, 1H, $J_{3,4} = 5.3$ Hz, $J_{4,5} = 11.9$ Hz, H-4c), 4.30 (d, 1H, $J_{1,2} = 7.8$ Hz, H-1a), 4.33 (m, 1H, H-2c), 4.47 (d, 1H, $J_{1,2} = 7.3$ Hz, H-1b), 4.80 (m, 1H, H-4d), 4.95 (m, 1H, H-4e), 5.06 (dd, 1H, $J_{2,3} = 8.9$ Hz, $J_{3,4} = 5.3$ Hz, H-3c), 5.22 (d, 1H, $J_{1,2} = 8.7$ Hz, H-1c), 5.27 (dd, 1H, $J_{6,7} = 2.5$ Hz, $J_{7,8} = 8.2$ Hz, H-7e), 5.3 (m, 2H, H-7d and H-8d), 5.44 (m, 1H, H-8e), and 6.99–7.84 (m, 29H, 6Ph).

Anal. Calcd for C₁₀₉H₁₃₅N₄₁O₁₉Si (2171.35): C, 60.29; H, 6.27; N, 1.94. Found: C, 60.01; H, 6.06; N, 1.85.

2-(Trimethylsilyl)ethyl O-(5-acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyranosylonic acid)-(2 \rightarrow 6)-O-(2-deoxy-3,4-O-isopropylidene-2-phthalimido- β -D-galactopyranosyl)-(1 \rightarrow 4)-O-[(5-acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyranosylonic acid)-(2 \rightarrow 3)]-O-(2,6-di-O-benzyl- β -D-galactopyranosyl)-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- β -D-glucopyranoside (**12**)

To a solution of **11** (600 mg, 0.28 mmol) in pyridine (15 mL) was added lithium iodide (384 mg, 2.96 mmol), and the mixture was stirred for 24 h under reflux and nitrogen atmosphere in the dark. The mixture was concentrated, and a solution of the residue in chloroform was successively washed with 2M HCl and water, dried (Na₂SO₄) and concentrated. Column chromatography (10:1 chloroform-methanol) of the residue on silica gel gave **12** (500 mg, 85%) as an amorphous mass: $[\alpha]_D -3.70^\circ$ (*c* 0.6, CHCl₃); the complete cleavage of the methyl ester was confirmed by ¹H-NMR.

Anal. Calcd for C₁₀₇H₁₃₁N₃O₄₁Si (2143.29): C, 59.96; H, 6.16; N, 1.96. Found: C, 59.85; H, 5.92; N, 1.87.

2-(Trimethylsilyl)ethyl O-(methyl 5-acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyranosylonate)-(2 \rightarrow 6)-O-(2-acetamido-2-deoxy- β -D-galactopyranosyl)-(1 \rightarrow 4)-O-[(methyl 5-acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyranosylonate)-(2 \rightarrow 3)]-O-(2,6-di-O-benzyl- β -D-galactopyranosyl)-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- β -D-glucopyranoside (**13**)

A solution of **3** (450 mg, 0.22 mmol) in aqueous 80% acetic acid (10 mL) was heated, with stirring, for 3 h at 40°C, then concentrated. Column chromatography (20:1 toluene-methanol) of the residue on silica gel gave **13** (375 mg, 85%) as an amorphous mass: $[\alpha]_D -7.18^\circ$ (*c* 0.6, CHCl₃); IR (KBr) 3600–3100 (OH and NH), 1760 and 1220 (ester), 1680 and 1560 (amide), 860 and 840 (Me₃Si), and 710 cm⁻¹ (phenyl); ¹H-NMR (CDCl₃) δ 1.03 (m, 2H, Me₃SiCH₂CH₂), 1.70–2.12 (11s, 33H, AcN and 10AcO), 1.84 (t, 1H, *J*_{gem} = *J*_{3ax,4} = 12.6 Hz, H-3e_{ax}), 2.11 (m, 1H, H-3d_{ax}), 2.21 (m, 1H, H-3d_{eq}), 2.60 (dd, 1H, *J*_{gem} = 12.6 Hz, *J*_{3eq,4} = 4.6 Hz, H-3e_{eq}), 3.80 and 3.84 (2s, 6H, 2MeO), 4.83 (m, 1H, H-4e), 5.11 (m, 1H, H-4d), and 7.20–7.46 (m, 25H, 5Ph).

Anal. Calcd for C₁₀₀H₁₃₁N₃O₄₀Si (2043.22): C, 58.78; H, 6.46; N, 2.06. Found: C, 58.72; H, 6.19; N, 1.98.

2-(Trimethylsilyl)ethyl O-(methyl 5-acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyranosylonate)-(2 \rightarrow 3)-O-(2,4,6-tri-O-benzoyl- β -D-galactopyranosyl)-(1 \rightarrow 3)-O-[(methyl 5-acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyranosylonate)-(2 \rightarrow 6)]-O-(2-acetamido-2-deoxy- β -D-galactopyranosyl)-(1 \rightarrow 4)-O-[(methyl 5-acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyranosylonate)-(2 \rightarrow 3)]-O-(2,6-di-O-benzyl- β -D-galactopyranosyl)-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- β -D-glucopyranoside (**15**)

To a solution of **13** (500 mg, 0.24 mmol) and methyl O-(methyl 5-acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyranosylonate)-(2 \rightarrow 3)-2,4,6-tri-O-benzoyl-1-thio- β -D-galactopyranoside [**33**] (**14**; 488 mg, 0.48 mmol) in dichloromethane (6 mL) was added molecular sieves 4 Å (660 mg), and the mixture was stirred for 6 h at room temperature then cooled to 0°C. DMTST [22,25] (504 mg, 1.2 mmol), was added to the mixture, and the stirring was continued for 24 h at 0°C. The precipitates were removed by filtration, and washed thoroughly with chloroform. The combined filtrate and washings was successively washed with M Na₂CO₃ and water, dried (Na₂SO₄) and concentrated. Column chromatography (15:1 toluene-methanol) of the residue on silica gel gave **15** (696 mg, 95%) as an amorphous mass: $[\alpha]_D +0.43^\circ$ (*c* 0.6, CHCl₃); IR (KBr) 3600–3100 (OH and NH), 1740 and 1220 (ester), 1660 and 1530 (amide), 860 and 840 (Me₃Si), 750 and 700 cm⁻¹ (phenyl); ¹H-NMR (CDCl₃) δ 1.02 (m, 2H, Me₃SiCH₂CH₂), 1.55–2.13 (16s, 48H, 4AcN and 12AcO), 1.59 (t, 1H, *J*_{gem} = *J*_{3ax,4} = 12.6 Hz, H-3e_{ax}), 1.84 (m, 1H, H-3e_{ax}), 2.05 (m, 1H, H-3f_{ax}), 2.42 (dd, 1H, *J*_{gem} = 12.6 Hz, *J*_{3eq,4} = 4.6 Hz, H-3e_{eq}), 2.54 (m, 2H, H-3e_{eq} and H-3f_{eq}), 3.71, 3.77, and 3.81 (3s, 9H, 3MeO), 4.80 (m, 1H, H-4g), 4.83 (m, 1H, H-4c), 4.97 (m, 1H, H-4f), 5.04 (d, 1H, *J*_{1,2} = 8.0 Hz, H-1d), 5.43 (d, 1H, *J*_{3,4} = 3.2 Hz, H-4d), 5.51 (dd, 1H, *J*_{1,2} = 8.0 Hz, *J*_{2,3} = 10.3 Hz, H-2d), and 7.17–8.16 (m, 40H, 8 Ph).

Anal. Calcd for C₁₄₇H₁₈₀N₄O₆₀Si (2991.12): C, 59.03; H, 6.07; N, 1.87. Found: C, 58.77; H, 5.78; N, 1.84.

2-(Trimethylsilyl)ethyl O-(methyl 5-acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyranosylonate)-(2 \rightarrow 3)-O-(2,4,6-tri-O-benzoyl- β -D-galactopyranosyl)-(1 \rightarrow 3)-O-[(methyl 5-acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyranosylonate)-(2 \rightarrow 6)]-O-(2-acetamido-4-O-acetyl-2-deoxy- β -D-galactopyranosyl)-(1 \rightarrow 4)-O-[(methyl 5-acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyranosylonate)-(2 \rightarrow 3)]-O-(2,6-di-O-acetyl- β -D-galactopyranosyl)-(1 \rightarrow 4)-2,3,6-tri-O-acetyl- β -D-glucopyranoside (**16**)

A solution of **15** (600 mg, 0.20 mmol) in ethanol (15 mL) was hydrogenated in the presence of 20% Pd(OH)₂ (600 mg) for 72 h at room temperature, the catalyst removed by filtration and the solution concentrated. The residue was treated with acetic anhydride (3 mL) and pyridine (6 mL) for 24 h at 35°C.

The mixture was concentrated, and a solution of the residue in chloroform was successively washed with 2M HCl and M Na₂CO₃, dried (Na₂SO₄) and concentrated. Column chromatography (15:1 chloroform–methanol) of the residue on silica gel gave **16** (563 mg, quantitative) as an amorphous mass: $[\alpha]_D -12.50^\circ$ (*c* 0.6, CHCl₃); IR (KBr) 3300 (NH), 1740 and 1220 (ester), 1670 and 1550 (amide), 860 and 840 (Me₃Si), and 710 cm⁻¹ (phenyl); ¹H-NMR (CDCl₃) δ 0.92 (m, 2H, Me₃SiCH₂CH₂), 1.53–2.20 (22s, 66H, 4AcN and 18AcO), 1.59 (t, 1H, $J_{\text{gem}} = J_{3\text{ax},4} = 12.2$ Hz, H-3gax), 1.71 (t, 1H, $J_{\text{gem}} = J_{3\text{ax},4} = 12.4$ Hz, H-3eax), 1.90 (m, 1H, H-3fax), 2.45 (m, 2H, H-3feq, and H-3geq), 2.85 (dd, 1H, $J_{\text{gem}} = 12.4$ Hz, $J_{3\text{eq},4} = 4.3$ Hz, H-3eeq), 3.76, 3.78, and 3.81 (3s, 9H, 3MeO), 4.98 (d, 1H, $J_{1,2} = 7.6$ Hz, H-1d), 5.40 (d, 1H, $J_{3,4} = 3.7$ Hz, H-4d), and 7.33–8.18 (m, 15H, 3Ph).

Anal. Calcd for C₁₂₄H₁₆₂N₄O₆₆Si (2792.72): C, 53.33; H, 5.85; N, 2.01. Found: C, 54.96; H, 5.87; N, 1.88.

O-(Methyl 5-acetamido-4,7,8,9-tetra-*O*-acetyl-3,5-dideoxy-*D*-glycero- α -*D*-galacto-2-nonulopyranosylonate)-(2→3)-*O*-(2,4,6-tri-*O*-benzoyl- β -*D*-galactopyranosyl)-(1→3)-*O*-[(methyl 5-acetamido-4,7,8,9-tetra-*O*-acetyl-3,5-dideoxy-*D*-glycero- α -*D*-galacto-2-nonulopyranosylonate)-(2→6)]-*O*-(2-acetamido-4-*O*-acetyl-2-deoxy- β -*D*-galactopyranosyl)-(1→4)-*O*-[(methyl 5-acetamido-4,7,8,9-tetra-*O*-acetyl-3,5-dideoxy-*D*-glycero- α -*D*-galacto-2-nonulopyranosylonate)-(2→3)]-*O*-(2,6-di-*O*-acetyl- β -*D*-galactopyranosyl)-(1→4)-2,3,6-tri-*O*-acetyl-*D*-glucopyranose (**17**)

To a solution of **16** (170 mg, 0.060 mmol) in dichloromethane (1 mL) was added trifluoroacetic acid (2 mL) at 0°C, and the mixture was stirred for 3 h at room temperature and concentrated. Column chromatography (15:1 chloroform–methanol) of the residue on silica gel gave **17** (150 mg, 92%) as an amorphous mass: IR (KBr) 3600–3100 (OH, NH), 1740 and 1220 (ester), 1670 and 1550 (amide), and 710 cm⁻¹ (Ph).

Anal. Calcd for C₁₁₉H₁₅₀N₄O₆₆ (2692.48): C, 55.09; H, 5.62; N, 2.08. Found: C, 52.89; H, 5.46; N, 1.85.

O-(Methyl 5-acetamido-4,7,8,9-tetra-*O*-acetyl-3,5-dideoxy-*D*-glycero- α -*D*-galacto-2-nonulopyranosylonate)-(2→3)-*O*-(2,4,6-tri-*O*-benzoyl- β -*D*-galactopyranosyl)-(1→3)-*O*-[(methyl 5-acetamido-4,7,8,9-tetra-*O*-acetyl-3,5-dideoxy-*D*-glycero- α -*D*-galacto-2-nonulopyranosylonate)-(2→6)]-*O*-(2-acetamido-4-*O*-acetyl-2-deoxy- β -*D*-galactopyranosyl)-(1→4)-*O*-[(methyl 5-acetamido-4,7,8,9-tetra-*O*-acetyl-3,5-dideoxy-*D*-glycero- α -*D*-galacto-2-nonulopyranosylonate)-(2→3)]-*O*-(2,6-di-*O*-acetyl- β -*D*-galactopyranosyl)-(1→4)-2,3,6-tri-*O*-acetyl-*D*-glucopyranosyl trichloroacetimidate (**18**)

Treatment of **17** (480 mg, 0.18 mmol) with trichloroacetonitrile (780 μ L, 5.4 mmol) and DBU (32 μ L, 0.22 mmol) in

dichloromethane (2 mL), then work up as described for **9** followed by column chromatography (20:1 chloroform–methanol) of the residue on silica gel gave **18** (505 mg, quantitative) as an amorphous mass: $[\alpha]_D -1.4^\circ$ (*c* 0.6, CHCl₃); IR (KBr) 3200 (NH), 1740 and 1220 (ester), 1650 and 1540 (amide), and 710 cm⁻¹ (phenyl); ¹H-NMR for α isomer (CDCl₃) δ 1.53–2.19 (22s, 66H, 4AcN and 18AcO), 2.45 (m, 2H, H-3feq, and H-3geq), 2.87 (dd, 1H, $J_{\text{gem}} = 13.0$ Hz, $J_{3\text{eq},4} = 4.3$ Hz, H-3eeq), 3.77, 3.79, and 3.81 (3s, 9H, 3MeO), 3.87 (t, 1H, $J_{3,4} = J_{4,5} = 9.8$ Hz, H-4a), 5.00 (d, 1H, $J_{1,2} = 7.8$ Hz, H-1d), 5.05 (dd, 1H, $J_{1,2} = 3.9$ Hz, $J_{2,3} = 10.3$ Hz, H-2a), 5.47 (t, 1H, $J_{2,3} = J_{3,4} = 9.8$ Hz, H-3a), 6.48 (d, 1H, $J_{1,2} = 3.9$ Hz, H-1a), 7.39–8.18 (m, 15H, 3Ph), and 8.65 (s, 1H, C=NH).

Anal. Calcd for C₁₂₁H₁₅₀Cl₃N₄O₆₆ (2836.87): C, 51.23; H, 5.33; N, 2.47. Found: C, 51.03; H, 5.14; N, 2.25.

O-(Methyl 5-acetamido-4,7,8,9-tetra-*O*-acetyl-3,5-dideoxy-*D*-glycero- α -*D*-galacto-2-nonulopyranosylonate)-(2→3)-*O*-(2,4,6-tri-*O*-benzoyl- β -*D*-galactopyranosyl)-(1→3)-*O*-[(methyl 5-acetamido-4,7,8,9-tetra-*O*-acetyl-3,5-dideoxy-*D*-glycero- α -*D*-galacto-2-nonulopyranosylonate)-(2→6)]-*O*-(2-acetamido-4-*O*-acetyl-2-deoxy- β -*D*-galactopyranosyl)-(1→4)-*O*-[(methyl 5-acetamido-4,7,8,9-tetra-*O*-acetyl-3,5-dideoxy-*D*-glycero- α -*D*-galacto-2-nonulopyranosylonate)-(2→3)]-*O*-(2,6-di-*O*-acetyl- β -*D*-galactopyranosyl)-(1→4)-2,3,6-tri-*O*-acetyl- β -*D*-glucopyranosyl)-(1→1)-(2*S*,3*R*,4*E*)-2-azido-3-*O*-(butyldiphenylsilyl)-4-octadecene-1,3-diol (**20**)

To a solution of **18** (100 mg, 0.035 mmol) and (2*S*,3*R*,4*E*)-2-azido-3-*O*-(butyldiphenylsilyl)-4-octadecene-1,3-diol [**19**; 40 mg, 0.070 mmol] in dichloromethane (1.5 mL) were added molecular sieves 4 Å (AW-300; 1.2 g), and the mixture was stirred for 12 h at room temperature, then cooled to 0°C. TMSOTf (1.4 μ L, 7.0 μ mol) was added, and the mixture was stirred for 45 h at 0°C, then filtered. The insoluble materials were washed with chloroform, and the combined filtrate and washings was washed with M NaHCO₃ and water, dried (Na₂SO₄) and concentrated. Column chromatography (20:1 chloroform–methanol) of the residue on silica gel gave **20** (71 mg, 62%) as an amorphous mass: $[\alpha]_D -14.25^\circ$ (*c* 0.8, CHCl₃); IR (KBr) 3300 (NH), 3100–2900 (CH), 2100 (azide), 1740 and 1220 (ester), 1670 and 1540 (amide), 710 and 700 cm⁻¹ (phenyl); ¹H-NMR (CDCl₃) δ 0.88 (t, 3H, $J_{\text{Me,CH}_2} = 6.6$ Hz, MeCH₂), 1.04 (s, 9H, Me₃C), 1.27 (s, 22H, 11CH₂), 1.73–2.22 (22s, 66H, 4AcN and 18AcO), 2.45 (m, 2H, H-3feq, and H-3geq), 2.85 (m, 1H, H-3eeq), 3.73, 3.77 and 3.80 (3s, 9H, 3MeO), 5.85 (m, 1H, H-5 of sphingosine), and 7.30–8.21 (m, 25H, 5Ph).

Anal. Calcd for C₁₅₃H₂₀₁N₇O₆₆Si (3238.36): C, 56.75; H, 6.26; N, 3.03. Found: C, 56.54; H, 6.11; N, 2.87.

O-(Methyl 5-acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyranosylate)-(2 \rightarrow 3)-O-(2,4,6-tri-O-benzoyl- β -D-galactopyranosyl)-(1 \rightarrow 3)-O-[(methyl 5-acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyranosylate)-(2 \rightarrow 6)]-O-(2-acetamido-4-O-acetyl-2-deoxy- β -D-galactopyranosyl)-(1 \rightarrow 4)-O-[(methyl 5-acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyranosylate)-(2 \rightarrow 3)]-O-(2,6-di-O-acetyl- β -D-galactopyranosyl)-(1 \rightarrow 4)-(2,3,6-tri-O-acetyl- β -D-glucopyranosyl)-(1 \rightarrow 1)-(2*S*,3*R*,4*E*)-3-O-(butyldi-phenylsilyl)-2-octadecanamido-4-octadecene-1,3-diol (**21**)

To a solution of **20** (83 mg, 0.025 mmol) in benzene (1 mL) and water (0.04 mL) was added triphenylphosphine (14 mg, 0.053 mmol), and the mixture was stirred for 24 h at 30°C and concentrated. To a solution of the residue in dichloromethane (1 mL) were added octadecanoic acid (22 mg, 0.077 mmol) and 1-ethyl-3-(3-dimethylaminopropyl)carbodiimide hydrochloride (16 mg, 0.083 mmol), and the mixture was stirred for 24 h at 30°C. The mixture was diluted with chloroform and the solution was washed with water, dried (Na₂SO₄) and concentrated. Column chromatography (20:1 chloroform-methanol) of the residue on silica gel gave **21** (68 mg, 76%) as an amorphous mass: $[\alpha]_D -2.20^\circ$ (*c* 1.2, CHCl₃); IR (KBr) 3200 (NH), 3100–2950 (CH), 1740 and 1240 (ester), 1650 and 1540 (amide), 710 and 700 cm⁻¹ (phenyl); ¹H-NMR (CDCl₃) δ 0.88 (t, 6H, *J*_{Me,CH₂} = 6.9 Hz, 2*MeCH*₂), 1.03 (s, 9H, *Me*₃C), 1.26 (s, 52H, 26*CH*₂), 1.77–2.18 (22s, 66H, 4AcN and 18AcO), 2.37–2.75 (m, 3H, H-3*eeq*, H-3*feq*, and H-3*geq*), 3.71, 3.77 and 3.81 (3s, 9H, 3*MeO*), 5.90 (m, 1H, H-5 of ceramide), and 7.36–8.25 (m, 25H, 5Ph).

Anal. Calcd for C₁₇₁H₂₃₇N₅O₆₈Si (3478.83): C, 59.04; H, 6.87; N, 2.01. Found: C, 58.91; H, 6.64; N, 1.75.

Ganglioside GT1ax

To a solution of **21** (18 mg, 0.019 mmol) in acetonitrile (1.5 mL) was added M tetrabutylammonium fluoride in tetrahydrofuran (0.3 mL), and the mixture was stirred for 48 h at room temperature, then concentrated. To a solution of the residue in methanol (1 mL) was added a catalytic amount of sodium methoxide, and the mixture was stirred for 24 h at room temperature. Water (0.5 mL) was added and the solution was stirred for 24 h at room temperature, then neutralized with Amberlite IR-120 (H⁺) resin. The resin was filtered off and washed with 1:1 chloroform-methanol, and the combined filtrate and washings was concentrated. Column chromatography (5:5:1 chloroform-methanol-water) of the residue on Sephadex LH-20 gave the ganglioside GT1ax (8.8 mg, 84%) as an amorphous mass: $[\alpha]_D +2.0^\circ$ (*c* 0.6, 5:5:1 CHCl₃-MeOH-H₂O); ¹H-NMR ((CD₃)₂SO-D₂O) δ 0.88 (t, 6H, *J*_{Me,CH₂} = 6.9 Hz, 2*MeCH*₂), 1.24 (s, 52H, 26*CH*₂), 2.00–2.02 (4s, 12H, 4AcN), 2.73–2.89 (m, 3H, H-3*eeq*, H-3*feq*, and H-3*geq*), 4.15 (d, 1H, *J*_{1,2} = 8.7 Hz, H-1a), 4.30 (d, 1H, *J*_{1,2} = 8.7 Hz, H-1b), 4.43 (d, 1H, *J*_{1,2} = 8.7 Hz, H-1d), 4.84 (m, 1H, H-1c), 5.32 (m, 1H, H-4 of ceramide), and 5.75 (m,

1H, H-5 of ceramide); FAB MS (negative ion mode, triethanolamine matrix): 2172.04 [M-Na]⁻ (C₉₅H₁₆₂N₅Na₂O₄₇ MW, Exact 2171.0236, Ave. 2172.3161), 2150.06, 2149.07 [M-2Na]⁻ (C₉₅H₁₆₃N₅NaO₄₇ MW, Exact 2149.0416, Ave. 2150.3343), 2127.06 [M-3Na]⁻ (C₉₅H₁₆₄N₅O₄₇ MW, Exact 2127.0597, Ave. 2128.3525), 1858.0, 1857.0 [M-2Na-NeuAc]⁻; 1696.0 [M-2Na-NeuAc-Gal]⁻, 1545.0 [M-3Na-2NeuAc]⁻, 888.7 [lactosylceramide]⁻, 726.6 [glucosylceramide]⁻, and 564.6 [Ceramide]⁻.

TLC-immunostaining of gangliosides

TLC of the gangliosides was carried out with a solvent system of chloroform/methanol/0.2% CaCl₂ (40:40:11) using an HPTLC plate (E. Merk, Darmstadt, Germany). GT1ax isolated from bovine brain was developed in parallel with ganglioside. Immunostaining of samples were performed by the methods previously described [41]. The developed plate was dried in vacuo for 20 min, and dipped in 0.4% polyisobutylmethacrylate in a solution of chloroform/*n*-hexane (16:84) for 1 min. Organic solvents were removed from the plate. The plate was covered with anti-Chol-1 α antibody (GGR41, ascites form) [42], and was incubated at room temperature overnight. The antibody was diluted with Tris-buffered saline (pH 7.4) (TBS) containing 0.3% gelatin. The plate was washed with TBS, and then covered with horseradish peroxidase-conjugated anti-mouse IgG. An enzyme-linked second antibody was diluted with TBS containing 0.3% gelatin and 5% skim milk. The plate was incubated at room temperature for 90 min, and then was washed with TBS. For visualization, the plate was incubated with ECL Western blotting detection reagents (Amersham, Buckinghamshire, England). Chemiluminescence was recorded on an X-ray film.

Acknowledgments

This work was supported by the Ministry of Education, Science and Culture of Japan (grant Nos. 09240101, 10556026 and 10660107). The authors are grateful to Dr. Takao Ikami of Sanwa Kagaku Kenkyusho for the FAB MS analyses.

References

- 1 Sawada N, Ishida H, Collins BE, Schnaar RL, Kiso M (1999) *Carbohydr Res* **316**: 1–5.
- 2 Hakomori S (1981) *Annu Rev Biochem* **50**: 733–64.
- 3 Ando S (1983) *Neurochem Int* **5**: 507–37.
- 4 Wiegandt H (1985) Gangliosides. In *Glycolipids—New Comprehensive Biochemistry*, Vol. 10, (Wiegandt H, ed) pp. 199–260. Amsterdam: Elsevier.
- 5 Ledeen RW, Hakomori S, Yates AJ, Schneider JS, Yu RK, eds (1998) *Ann NY Acad Sci* Vol. 845, Sphingolipids as signaling modulators in the nervous system: New York. The New York Academy of Sciences.

- 6 Ando S, Hirabayashi Y, Kon K, Inagaki F, Tate S, Whittaker VP (1992) *J Biochem* **111**: 287–90.
- 7 Hirabayashi Y, Nakao T, Irie F, Whittaker VP, Kon K, Ando S (1992) *J Biol Chem* **267**: 12973–8.
- 8 Derrington EA, Whittaker VP (1993) *Neuro Report* **4**: 317–19.
- 9 Borroni E, Derrington EA, Whittaker VP (1993) *Dev Brain Res* **71**: 247–52.
- 10 Irie F, Hidari K, Tai T, Li Y-T, Sayama Y, Hirabayashi Y (1994) *FEBS Lett* **351**: 291–4.
- 11 Irie F, Hashikawa T, Tai T, Sayama Y, Hirabayashi Y (1994) *Brain Res* **665**: 161–6.
- 12 Hotta K, Komba S, Ishida H, Kiso M, Hasegawa A (1994) *J Carbohydr Chem* **13**: 665–77.
- 13 Prabhanjan H, Aoyama K, Kiso M, Hasegawa A (1992) *Carbohydr Res* **233**: 87–99.
- 14 Hotta K, Ishida H, Kiso M, Hasegawa A (1995) *J Carbohydr Chem* **14**: 491–506.
- 15 Yang LJ-S., Zeller CB, Shaper NL, Kiso M, Hasegawa A, Shapiro RE, Schnaar RL (1996) *Proc Natl Acad Sci USA* **93**: 814–18.
- 16 Collins BE, Yang LJ-S, Mukhopadhyay G, Filbin MT, Kiso M, Hasegawa A, Schnaar RL (1997) *J Biol Chem* **272**: 1248–55.
- 17 Collins BE, Kiso M, Hasegawa A, Tropak MB, Roder JC, Crocker PR, Schnaar RL (1997) *J Biol Chem* **272**: 16889–95.
- 18 Kelm S, Pelz A, Schauer R, Filbin MT, Tong S, Bellard de M-E, Schnaar RL, Mahoney JA, Hartnell A, Bradfield P, Crocker PR (1994) *Curr Biol* **4**: 965–72.
- 19 Powell LD, Varki A (1995) *J Biol Chem* **270**: 14243–6.
- 20 Crocker PR, Clark EA, Filbin M, Gordon S, Jones Y, Kehrl JH, Kelm S, Douarin LcN, Powell L, Roder J, Schnaar RL, Sgroi DC, Stamenkovic I, Schauer R, Schachner M, van den Berg TK, van der Merwe PA, Watt SM, Varki A (1998) *Glycobiology* **8**: no. 2, V.
- 21 Ito H, Ishida H, Kiso M, Hasegawa A (1997) *Carbohydr Res* **304**: 187–189. Erratum: *ibid* (1998) 306: 579–85.
- 22 Kiso M, Hasegawa A (1994) *Methods Enzymol* **242**: 173–82.
- 23 Veeneman GH, van Leeuwen SH, van Boom JH (1990) *Tetrahedron Lett* **31**: 1331–4.
- 24 Konradsson P, Udodong UE, Fraser-Reid B (1990) *Tetrahedron Lett* **31**: 4313–16.
- 25 Fügedi P, Garegg PJ (1986) *Carbohydr Res* **149**: c9–c12.
- 26 Kanie O, Kiso M, Hasegawa A (1988) *J Carbohydr Chem* **7**: 501–6.
- 27 Murase T, Ishida H, Kiso M, Hasegawa A (1988) *Carbohydr Res* **184**: c1–c4.
- 28 Hasegawa A, Nagahama T, Ohki H, Kiso M (1992) *J Carbohydr Chem* **11**: 699–714.
- 29 Hasegawa A, Kiso M (1996) Chemical synthesis of sialyl glycosides. In *Preparative carbohydrate chemistry* (Hanessian S, ed) pp 357–79. New York: Dekker.
- 30 Marra A, Sinaÿ P (1990) *Carbohydr Res* **195**: 303–08.
- 31 Hasegawa A, Ohki H, Nagahama T, Ishida H, Kiso M (1991) *Carbohydr Res* **212**: 277–81.
- 32 Lemieux RU, Takeda T, Chung B (1976) *ACS Symp Ser* **39**: 90–115.
- 33 Kameyama A, Ishida H, Kiso M, Hasegawa A (1990) *Carbohydr Res* **200**: 269–85.
- 34 Jansson K, Ahlfors S, Frejd T, Kihlberg J, Magnusson G, Dahmen J, Noori G, Stenvall K (1988) *J Org Chem* **53**: 5629–47.
- 35 Magnusson G (1992) *Trends in Glycosci Glycotech* **4**: 358–67.
- 36 Mori M, Ito Y, Ogawa T (1990) *Carbohydr Res* **195**: 199–224.
- 37 Kiso M, Nakamura A, Tomita Y, Hasegawa A (1986) *Carbohydr Res* **158**: 101–11.
- 38 Ehara T, Kameyama A, Yamada Y, Ishida H, Kiso M, Hasegawa A (1996) *Carbohydr Res* **281**: 237–52.
- 39 Schnaar RL, Collins BE, Wright LP, Kiso M, Tropak MB, Roder JC, Crocker PR (1998) *Ann NY Acad Sci* **845**: 92–105.
- 40 Collins BE, Ito H, Ishida H, Kiso M, Schnaar RL (1998) *19th Int. Carbohydr Symp Abstr*, San Diego: CK005.
- 41 Marquina G, Waki H, Fernandez E.L, Kon K, Carr A, Variante O, Perez R, Ando S, (1996) *Cancer Res* **56**: 5165–71.
- 42 Kusumoki S, Chiba A, Hirabayashi Y, Irie F, Kotani M, Kawashima I, Tai T, Nagai Y (1993) *Brain Res* **623**: 83–88.

Received 18 June 1999, revised and accepted 19 November 1999